

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
21 February 2002 (21.02.2002)

PCT

(10) International Publication Number
WO 02/14868 A1

(51) International Patent Classification⁷: G01N 33/543

(74) Agents: NATARAJ, Guruswamy et al.; Subramaniam, Nataraj & Associates, E 556 Greater Kailash II, New Delhi 110 048, Maharashtra (IN).

(21) International Application Number: PCT/IN00/00075

(22) International Filing Date: 16 August 2000 (16.08.2000)

(81) Designated States (*national*): AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(25) Filing Language: English

(26) Publication Language: English

(71) Applicant (*for all designated States except US*): COUNCIL OF SCIENTIFIC AND INDUSTRIAL RESEARCH [IN/IN]; Rafi Marg New Delhi 110 001, Maharashtra (IN).

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): NAHAR, Pradeep [IN/IN]; Centre for Biochemical Technology, Mall Road, Delhi 110 007, Maharashtra (IN). BORA, Utpal [IN/IN]; Centre for Biochemical Technology, Mall Road, Delhi 110 007, Maharashtra (IN). SHARMA, Gaiinda, Lal [IN/IN]; Centre for Biochemical Technology, Mall Road, Delhi 110 007, Maharashtra (IN).

Published:

— with international search report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 02/14868 A1

(54) Title: A RAPID METHOD FOR MICROWAVE MEDIATED ENZYME-LINKED IMMUNOSORBENT ASSAY

(57) Abstract: This invention relates to a rapid and efficient method for carrying out enzyme-linked immunosorbent assay for detection of minute quantities of biomolecules such as antigen, antibody etc. This invention particularly relates to microwave mediated immobilization of antigen or antibody on to the activated surface followed by performing subsequent steps of ELISA by controlled microwave irradiation. The invented procedure has dramatically reduced the total time required for ELISA to less than 10 minutes from hours to days. The invented ELISA procedure is rapid, economical, reproducible and simple and can be automated. The invented procedure is useful for carrying out ELISA in clinical diagnostics, molecular biology, agriculture, sericulture, food technology, environmental science, biomedical research and other related fields.

A RAPID METHOD FOR MICROWAVE MEDIATED ENZYME-LINKED IMMUNOSORBENT ASSAY

FIELD OF THE INVENTION

The invention relates to a rapid method for carrying out microwave mediated ELISA (MELISA). More particularly, this invention relates to a rapid and efficient method for
5 microwave mediated ELISA (MELISA) wherein all the major steps of ELISA can be performed under microwave irradiation in short time. This method is useful in clinical diagnostics, molecular biology, agriculture, food technology, environmental science etc.

The invented ELISA method is simple, time saving and obviates the time consuming cumbersome procedure. This method has the potential for automation.

10 This method has the advantage over the existing methods for ELISA which take long time usually ranging from several hours to 2 days whereas the invented method takes less than 10 minutes. It is particularly useful for disease diagnosis where quick results are required.

BACKGROUND OF THE INVENTION

15 Enzyme linked immunosorbent assay (ELISA) is a very sensitive technique used for semiquantitative or quantitative determination of the concentration of certain antigens and antibodies. ELISA has become a useful tool in disease diagnosis in both animals and plants. Apart from this, its other applications include screening of monoclonal antibodies during the course of their production (Douillard, J.Y. and Hoffman, T., 1983), pesticide residue
20 detection in crop produce (Van Emon, J.M. and Lopez-Avila, V., 1992) and environmental samples like soil and water (Linde, D.G. and Goh, K.S., 1995), detection of apoptosis in tissue culture etc. (Salgame, P. et al, 1996). Polystyrene microtitre plate is used universally for carrying out ELISA, as it is transparent, cheap, easily available, can be moulded to any desired shape and has a property of binding proteins through adsorption. Conventional
25 methods of ELISA are based on the immobilization of antigen or antibody onto the surface of

the wells of a polystyrene microtitre plate through adsorption. This is attributed to the non-covalent interaction between the biomolecule and the polystyrene surface.

However, adsorption is usually too inefficient a process to give good yields and doesn't always proceed in a dose dependent manner. To overcome the inefficiency of conventional methods covalent immobilization of biomolecules onto the microtitre plate has been carried out by many (Sato, A. et al, 1999). Covalent binding of immunogens to grafted plastic surfaces has also been reported (Larsson, P.H. et al, 1987). Despite this, the conventional ELISA method requires very long time varying from several hours to 2 days for completion. This is the main drawback of different ELISA methods, either based on adsorption or covalent binding. In case of medical urgency precious time is lost in diagnosis before the patient could be given medication. In agriculture, ELISA is useful for detecting pesticide residues in crop produce and environmental samples. Export and marketing of crop produce can be delayed as a consequence of this handicap in the ELISA method, which contributes to loss of valuable foreign exchange.

The applicants have developed a novel and unique method whereby ELISA can be carried out rapidly by the use of microwaves. Microwaves are known for accelerating immunohistochemistry for about a decade (Boon, M.E and Kok, L.P., 1992; Boon, M.E. et al, 1989; Boon, M.E. et al, PCT patent application WO 89/ 03038; Chiu, K.Y. and Chan, K.W., 1987; Hjerpe, A. et al, 1988).

Covalent immobilization of antigen or antibody on to a polystyrene surface by microwave irradiation has not been reported so far. In fact, all the major steps of ELISA by microwave irradiation in such a short time to detect minute quantities of antigen or antibody by measuring optical density was not known in the prior art.

However, there are attempts for doing one of the steps of ELISA by microwave irradiation (Hjerpe, A. et al, 1988) where polystyrene ELISA plates were coated first with

rabbit anti-carcinoembryonic antigen by incubating over night at 4°C, followed by incubation of the antigen (CEA). In the subsequent step i.e. after the addition of enzyme-linked antibodies the authors studied the effect of microwave irradiation on antigen-antibody reactions.

5 In another experiment by the same authors, ELISA plates were first coated with normal mouse serum by incubating over night at 4°C, followed by overnight incubation with non-labeled rabbit anti-mouse Ig. In the subsequent step they added mouse PAP (peroxidase-antiperoxidase) complexes and determined the effect of microwave irradiation on the reactivity constants of reactions of this last step.

10 In third experiment Hjerpe et.al first coated the plates with non-specific mouse serum followed by incubation with biotinylated horse-anti-mouse IgG. The plates were then used to study the effect of microwaves upon the subsequent binding of biotin-avidin complexes.

The reaction yields in all the above experiments were smaller in samples subjected to microwave irradiation as compared to those processed without microwave stimulation. The experiments showed that the microwaves caused a major loss of reactivity and the total yields were approximately 10% to 15% compared to conventional one that was carried out outside the microwave oven. According to authors, the diminished value in the microwave technique may be due to too high temperatures in the wells, despite the fact that a water load (a beaker of water to absorb excess microwave energy) and chilled bottom plate were taken as a precautionary measure.

20 In further ELISA experiment authors (Koh and Boon, 1992) used a fiberoptic thermometer to restrict the temperature below 40°C. Here also a water load of 200 ml tap water was taken. Besides, the fluid in the wells was stirred by slowly blowing air through the solution via thin plastic tips, which were inserted into the wells. In an experiment the authors carried out only two steps namely, antibody and conjugate binding steps by microwave

irradiation for 6 minutes at 150 watts each.

In another experiment, antibody, antigen and conjugate binding steps were carried out in 15, 30 and 30 minutes respectively using 45-50% microwave power in each step.

Remaining steps in both the experiments were carried out by conventional procedure.

5 But the ELISA values were found to be much less in the above experiments than the conventional method.

According to authors, the longer exposure time gave higher extinction value (ELISA value) but the time gain was not attractive. In fact there was no benefit when exposure time of 30 minutes or more were used. Too short exposure times led to extinction values which, were
10 rather low and could not be used.

The reported methods of ELISA by microwave exposure has several drawbacks such as (1) the results (ELISA value) were much less than the conventional procedure, (2) the time gain was not attractive. It may be possible to get comparable results (ELISA value) by doing the ELISA out side microwave oven in the same time, (3) procedure required water load, (4)
15 it required cooling system or chilled bottom plate, (5) it needed a stirring system in the well of the microtitre plate, (6) not all the steps were carried out by microwave energy and the reported ELISA procedure has little or no potential for automation.

The applicants in the present invention have overcome all the above drawbacks. In fact, like thermal energy microwave can also activate or inactivate a biomolecule. Without
20 proper conditions the microwave may cause partial or total destruction of the biomolecule leading to low or undesirable ELISA value. In the invented procedure proper conditions were found, most of, which are contrary to the reported method, or not known in the prior art. In the invented method all the steps except color development are carried out by microwave stimulation. Blocking step was not carried out by microwave irradiation in the published
25 procedure as it gave nonspecific binding, whereas the applicants have invented a method

where blocking step is carried out in short time by microwave irradiation. Longer microwave exposure time gave higher ELISA value in the reported method whereas in the invented procedure, it leads to destruction of material or nonspecific binding. This may be because water load or the cooling system used in the reported method absorbed most of the microwave energy giving minimum microwave effect and a significant effect of time as in the conventional method. In contrast, the invented method does not require any water load, cooling system or stirring system. Moreover, in the invented method about 200 times less (excluding color development step) time requires for ELISA than the conventional one (which takes around 18 hours) with comparable or even better ELISA value. Hence, it also has a great potential for automation.

The reported ELISA was carried out on a microtitre plate that did not bind the biomolecule through covalent linkage. In fact, it is known that the microwaves may readily effect the integrity of the noncovalent secondary bonding, such as hydrogen bonds, hydrophobic interactions and Van der Waal's interaction.

OBJECTS OF THE INVENTION

The main object of the present invention is to provide a rapid and efficient method for enzyme linked immunosorbent assay to detect minute quantities of antigen or antibody spectrophotometrically for rapid diagnosis of diseases.

Another object of the present invention is to provide a technique, which is simple, reproducible and does not require additional expertise or costly equipment.

Another object of the present invention is to provide a rapid technique, which has the potential for automation and which can minimize human error that usually varies from person to person.

SUMMARY OF THE INVENTION

With a view to achieve the objects and overcoming the disadvantage of known ELISA

method, a rapid and efficient method is provided for microwave mediated ELISA (MELISA) which comprises the steps of (i) covalent immobilization of the antigen or antibody on to the activated solid surface by microwave irradiation, (ii) blocking the free surface with blocking agent by brief microwave irradiation, (iii) binding of antibody or antigen by controlled
5 microwave irradiation (iv) binding of conjugate by controlled microwave irradiation, (v) adding dye-substrate to the wells and (vi) recording the absorbance value.

Microwave mediated ELISA (MELISA) procedure is carried out on an activated surface in a very short time (around 10 minutes) and with the same efficacy as in the conventional ELISA carried out at 37°C, in 16-18 hours.

10 This invention has the potential of automation or semi automation of the ELISA procedure.

Microwaves are known to effect the integrity of the noncovalent secondary bonding, such as hydrogen bonds, hydrophobic interactions and Van der Waal's interaction.

To overcome this problem the applicants activates the surface of the microtitre wells
15 prior to use. The activated surface immobilizes the antigen through a covalent bond by a brief microwave exposure. This covalently immobilized antigen is stable enough to withstand repeated but brief microwave exposure, which were needed for performing the subsequent steps of ELISA. Subsequent steps of biomolecule binding in the ELISA procedure are through non covalent binding which are susceptible to microwave energy. These problems
20 are overcome by controlling the time and energy of the microwave irradiation in each step.

Novelty of the present invention is that the ELISA is carried out on to an activated surface capable of forming covalent linkage with the proteinaceous ligand.

Microwave mediated covalent immobilization of biomolecule, more preferably antigen or antibody on to the activated surface is another novelty of this invention.

25 Controlled microwave irradiation in each step of ELISA procedure is another new

approach.

In the invented procedure, all the steps of ELISA such as antigen binding, blocking, antibody and conjugate binding are carried out by microwave irradiation and only enzyme-substrate reaction is performed outside microwave oven at room temperature. The invented
5 procedure of ELISA is very fast with comparable or even better ELISA value than the conventional method.

Another novelty of the present invention is that the invented ELISA procedure does not require any water load or stirring system.

Another novelty of the present invention is that the invented ELISA procedure can be
10 fully or partially automated with the use of a specially designed device.

Another novelty of the present invention is that the invented procedure can be used for other immunoassays like radio immunoassay, radio-immunosorbent test, radio allergosorbent test, biotin- avidin /streptavidin immunoassay, immunoblotting, immunostaining etc. apart from different types of ELISA such as direct ELISA, indirect
15 ELISA, sandwich ELISA and alike.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a new approach for enzyme linked immunosorbent assay technique on an activated microtitre plate, module or well by microwave exposure. Activated surface immobilizes the antigen through covalent bonding by microwave
20 irradiation. This covalently immobilized antigen is stable enough to withstand repeated but brief microwave exposure, which are needed for performing the subsequent steps of ELISA. ELISA is a multistep and delicate process where improper condition in any step may hamper the whole results.

In the invented procedure inert solid surface such as polystyrene is activated by
25 photochemical reaction in dry condition using a photoactivable compound. Activation of the

solid support is made by exposing the support coated with the photoactivable compound to UV radiation or bright sunlight. This activated support is used for microwave mediated ELISA (MELISA) and for control ELISA to detect antibodies, to example *E.histolytica* and *Aspergillus fumigatus*. When untreated support is used it failed to bind the biomolecule in
5 such a short time by microwave irradiation.

Microwave irradiation is performed inside a domestic microwave oven (BPL-Sanyo, India), operating at a frequency of around 2450 MHz.

The amoebic antigen is obtained from the culture of *E.histolytica* as per published procedure (Sawhney, S., et al, 1980; Sharma, G.L, et al, 1984). The protein concentration of
10 the antigens was 1.54 mg per ml as determined by the method of Lowry *et.al.* (Lowry, O.H., et al, 1951). The antigen was diluted before performing ELISA and a concentration of 1.0 μ g per well was used for coating the wells.

Phosphate buffer saline (PBS), pH 7.2, 0.01 M is used as coating buffer.

Phosphate buffer saline (PBS), pH 7.2, 0.01 M together with 0.1% tween 20 is used as
15 washing buffer.

Blocking solution is made by dissolving 2% BSA in 0.01 M PBS, pH 7.2.

Substrate solution is prepared by adding 0.067% of o-phenylenediamine and 0.043% of H₂O₂ in 0.1 M phosphate citrate buffer, pH 4.5).

Hyperimmune sera to *E.histolytica* is raised in Newzealand White rabbits as per the
20 methods of Sawhney *et al* (Sawhney *et. al*, 1980). The antibody titres in the sera are checked by gel-diffusion test.

The antibody (+ve sera) is diluted before performing ELISA and 1:300 dilution in PBS is used for the experiments.

Rabbit is bled through the ear vein before giving the immunization dose of
25 *E.histolytica* antigen to obtain negative control sera (-ve sera). 100 μ l of diluted (1:300) -ve

sera is used for each well.

Horse radish peroxidase conjugated anti-rabbit IgG is purchased from Sigma as lyophilized powder.

After reconstitution, the optimum dilution is found to be 1: 4000 as determined by
5 checkerboard titration, which is used in the experiments.

ELISA is a 5-step procedure, namely antigen binding, blocking, antibody binding, conjugate binding and color development. Each step of MELISA is optimized by carrying out the subsequent steps by conventional ELISA procedure. Conventional ELISA is carried out
10 by overnight coating the activated wells with antigen at 4°C, blocking the wells in 2 hours at 37°C followed by antibody and conjugate binding at 37°C for 2 h each and color development that is enzyme-substrate reaction at room temperature for 5 minutes followed by reading absorbance.

In the invented procedure, the first step of ELISA is performed by covalent immobilization of the amoebic antigen onto the activated polystyrene microwell plate by
15 microwave irradiation at 700 watts in different duration of time. Binding, is detectable even in 10 seconds which increases with the increase in time of irradiation (Table 1). At 90 seconds the antigen binding becomes more or less same as 70 seconds of microwave irradiation. Hence, optimum time for antigen immobilization is taken as 70 seconds. In control experiments carried out for the same duration that is 70 seconds at 37°C, no binding
20 of antigen to the activated polystyrene surface is observed.

In the second step of MELISA, blocking is carried out with 2% BSA in 10 seconds in the microwave oven at a power output of 700 watts to block the free surface of the activated surface. Further increase in irradiation time showed non-specific binding (Table 2.)

In the third step of MELISA, antibody binding to the immobilized antigen is carried
25 out at 155 watts for 100 seconds. This is a crucial step where harsh conditions such as excess

time or higher power output leads to nonspecific binding as shown in Table-3. At 155 watts in 50 seconds and 100 seconds, no nonspecific binding is observed. Although the observed OD is very low in 50 seconds, 100 seconds is found to be excellent, giving high ratio of -ve to +ve sera. Increase in time to 150 seconds increases the nonspecific binding (table 4).

5 In the fourth step of MELISA, conjugate binding is carried out by microwave irradiation at 155 watts for different duration of time. Excellent result is obtained in 100 seconds (Table 6). Harsh condition such as excess time or higher energy such as 700 watts leads to nonspecific binding (Table 5).

10 In each step control experiment is carried out by doing the same experiment outside microwave oven for the same duration of time. In all this experiments negligible or undesirable ELISA value (nonspecific binding) is obtained.

After optimizing each step of MELISA, the applicants carried out all the steps with optimized conditions of microwave irradiation without damaging the biomolecules. To achieve it the applicants have immobilized antigen on to the activated well in 70 seconds, 15 blocking in 10 seconds, antibody binding in 100 seconds and conjugate binding at 100 seconds of irradiation. The total time required for all these steps are 280 seconds only by the invented procedure whereas conventional ELISA is done in about 18 hours.

MELISA and ELISA to detect *E.histolytica* antibodies are repeated several times under similar experimental conditions and the results are found to be highly reproducible as 20 shown in Table- 7.

MELISA procedure is further verified by detecting *Aspergillus fumigatus* antibody from patients sera. *A. fumigatus* antigen is obtained from static culture as per published procedure (Banerjee, B., et al, 1990).

25 The protein concentration of the antigens is found to be 12.5 mg per ml as determined by the method of Lowry *et.al.* (Lowry *et.al.*, 1951). Immunoreactivity of the antigen is

checked by using hyperimmune serum raised in Newzealand White rabbits.

Sera samples are obtained from 10 patients of allergic bronchopulmonary aspergillosis (ABPA). All these patients fulfill the clinical criteria of ABPA as described earlier (Rosenberg, M., et al, 1997).

5 Negative control sera samples are taken from 10 volunteers who are apparently healthy and do not have any respiratory or other disease.

Pooled positive and negative sera are taken as the controls for ELISA, which are found to be comparable in both MELISA and conventional ELISA.

10 Horse radish peroxidase conjugated with anti-human IgG is purchased from Sigma as lyophilized powder. After reconstitution, the optimum dilution of the conjugate is found to be 1:4000 as determined by checkerboard titration.

A.fumigatus antibody (present in patient sera) detected by the invented method is in agreement with the conventional ELISA procedure (Table.8)

The results obtained in different experiments are given comparative grades as below:

15 Excellent = + + + +

Very good = + + +

Good = + +

Poor = +

Undesirable or no results = -

20 The total time required for MELISA is less than 10 minutes. However, time required in each step may vary depending on the biomolecules where excellent results can be obtained by minor modification of reaction conditions that is minor alternation in duration and energy of irradiation.

25 Accordingly the present invention provides a rapid method for microwave mediated enzyme-linked immunosorbent assay characterized in using an activated solid support

wherein the said method comprises:

(a) providing an activated solid support,

(b) loading a biomolecule selected from an antigen or antibody by dissolving the said biomolecule in a coating buffer into the activated well of the said solid support and
5 placing the said well inside a microwave oven followed by irradiating the said well with microwaves at a frequency ranging between 2300-2500 MHz with the power output ranging between 600-900 watts for a period ranging from 50-100 seconds followed by washing the well thoroughly with an appropriate washing buffer,

(c) blocking the free sites of the well with an immobilized biomolecule as obtained from step

10 (b) as above by loading blocking solution into the said well and irradiating it inside the microwave oven at a frequency of from 2300-2500 MHz with a power out put ranging between 600-800 watts for a period ranging from 5-20 seconds and washing the said well with an appropriate washing buffer,

(d) loading the corresponding antibody or antigen dissolved in a buffer into the well

15 immobilized with antigen or antibody as obtained from step (c) above followed by irradiation of said well inside the microwave oven at a frequency of from 2300-2500 MHz with a power output ranging from 50-200 watts for a period ranging from 90-200 seconds followed by washing with washing buffer,

(e) loading an appropriate enzyme- conjugate dissolved in a suitable buffer into the above

20 said well obtained from step (d) and irradiating the said well inside a microwave oven at a frequency of from 2300-2500 MHz with a power output ranging from 100-300 watts for a period ranging from 50-150 seconds followed by washing with a washing buffer,

(f) adding a substrate-dye-buffer to the above well as obtained from step (e) as above and keeping it for a period ranging from 4 to 10 minutes in dark followed by addition of stop

25 solution and measuring optical density of the solution by spectrophotometer at a suitable

wavelength.

In an embodiment of the present invention the solid support used is selected from the group consisting of materials such as polystyrene, polypropylene, polyethylene, glass, cellulose, nitrocellulose, silicagel, polyvinyl chloride, polyaniline and alike.

5 In an embodiment of the present invention the preferred solid support used is polystyrene.

In yet another embodiment the solid support is selected from any shape, form and size such as sheets, plates, test particles such as beads and microspheres, test tubes, test sticks, test strips, well, ELISA plate, microwell plate or module.

10 In an embodiment of the present invention the solid support used for immobilizing biomolecules is selected from any support having at least one active functional group capable of binding ligand molecules by covalent means.

In yet another embodiment of the present invention the functional group is selected from halide, aldehyde, acetyl, epoxide, succinamide, isothiocyanate, acylazide and alike.

15 In yet another embodiment of the present invention the functional group may be present in the support itself or can be introduced by conventional chemical or photochemical or other methods known to prior art.

In yet another embodiment of the present invention the functional group is introduced on to the solid support by photochemical reaction in dry condition using a photoactivable
20 compound which is selected from 4-fluoro-3-nitroazidobenzene, N-hydroxysulfo-succinimidyl 4-azidobenzoate, N-hydroxysulfo-succinimidyl 4-azidosalicylic acid and alike.

In yet another embodiment of the present invention polystyrene surface is activated by coating with 1-flouro-2-nitro-azidobenzene and exposing the coated support in dry condition to UV radiation at 365nm.

25 In yet another embodiment of the present invention the light source for photochemical

reaction is selected from UV lamp, laser beam, bright sunlight or alike.

In yet another embodiment of the present invention time for photoreaction for activation of solid support is selected from 10 seconds to 10 hours.

In yet another embodiment microwave irradiation is performed in a microwave apparatus selected from domestic microwave oven, specially designed microwave oven or
5 any apparatus or chamber in which microwave is generated and alike.

In yet another preferred embodiment of the invention, the first step of ELISA, is carried out by covalent binding of antigen or antibody onto the activated plate by microwave irradiation at a frequency of from 2300-2500 MHz with the power output ranging between
10 600-900 watts for a short period ranging from 50-100 seconds.

In yet another preferred embodiment of the invention the second step of ELISA, that is the blocking step is carried out by microwave irradiation at a microwave frequency of from 2300-2500 MHz with a power out put ranging between 600-800 watts for a short period ranging from 5-20.

15 In yet another preferred embodiment of the invention the third step of ELISA, that is corresponding antibody or antigen binding is carried out by microwave irradiation at the microwave frequency of from 2300 to 2500 MHz with power output of from 50 to 200 watts in a period ranging from 90 to 200 seconds.

In yet another preferred embodiment of the invention the fourth step of ELISA, that is
20 enzyme-conjugate binding is carried out by microwave irradiation at the microwave frequency of from 2300 to 2500 MHz with power output of from 100 to 300 watts in a period ranging from 50 to 150 seconds.

In yet another preferred embodiment of the invention the total time for antigen binding, blocking, antibody binding and conjugate binding is ranging from 195 to 470
25 seconds wherein the total time for conventional method usually is ranging from 10 hours to

24 hours.

In another embodiment to the present invention, the antigen can be dissolved in a coating buffer of suitable composition having a pH, in the range of from 6.5 to 11 with molarity ranging from 0.005 M to 0.1 M compatible with the antigen such as carbonate
5 buffer, phosphate buffer and alike.

In yet another preferred embodiment of the invention, washing buffer used is a mixture of phosphate buffer having a pH, in the range of from 6.5 to 11, with molarity ranging from 0.005 M to 0.1 M and tween 20 in the range between 0.05% to 3%.

In yet another preferred embodiment of the invention, blocking reagent is selected
10 from bovine serum albumin, skimmed milk powder, gelatin and alike.

In another embodiment to the present invention biomolecule is selected from antigen or antibody. Antigen may be any, biomolecule, microorganism, substance etc. that elicits or has the potential to elicit an immune response.

In yet another preferred embodiment of this invention, antibody is selected from any
15 biomolecule, which is produced by the host in response to inoculation with the specific antigen and has capabilities of binding to the antigen in a specific manner.

In yet another preferred embodiment of this invention, conjugate is a specific biomolecule having antibody or antigen conjugated with an enzyme selected from peroxidase or alkaline phosphatase.

20 In yet another preferred embodiment of this invention, enzyme may be replaced by a label selected from chromophore, fluorophore and alike which facilitates its assay.

In yet another preferred embodiment of this invention, the invented procedure can be used for other immunoassays like radio immunoassay, radio-immunosorbent test, radio allergosorbent test, biotin- avidin/streptavidin immunoassay, immunoblotting, immunostaining etc. apart
25 from different types of ELISA such as direct ELISA, indirect ELISA, sandwich ELISA and

alike.

The invention further provides an apparatus for microwave mediated enzyme linked immunosorbent assay (MELISA) comprising (a) a loading chamber, for loading the samples or reagents from a specified bottle from a fine tube by a suitable pump onto the activated polystyrene plate/module automatically; (b) a reaction chamber consisting of magnetron, exhaust fan etc. for carrying out all the steps such as binding of the antigen, blocking, antibody binding and antibody enzyme conjugate binding by microwave irradiation and enzyme substrate reaction without microwave stimulation at ambient temperature in a pre-programmed time; (c) a washing cum drying chamber for washing and drying the said ELISA plate or module automatically by a pre-programmed command after each step of the MELISA procedure; (d) a detection chamber for colorimetric detection with the help of the spectrophotometer; (e) a moving platform is used for carrying the Elisa plate/modules from one chamber to another chamber; (f) a microprocessor based computing means for controlling MELISA method through suitable hardware and software.

This invention is further explained with the help of the following examples and should not be construed to limit the scope of the invention.

EXAMPLE 1

Activation of solid support.

Wells of a module (12 well polystyrene module, Dynatech, USA) are loaded with 1.82 mg 1-flouro-2-nitro-azidobenzene (FNAB), dissolved in 100µl of methanol per well and dried properly in the dark. FNAB coated wells are then irradiated for 10 min. by UV light at 365 nm in a UV Stratalinker 2400 (Stratagene®, USA) or kept under bright sunlight for 1 h. The wells are then washed several times with methanol to remove the unbound linker and dried at room temperature. These activated wells of the module are used for immobilization of antigens or antibodies in the invented procedure.

EXAMPLE 2**Immobilization of *Entamoeba histolytica* antigen by microwave irradiation.**

E. histolytica antigen (1 μ g) diluted in 100 μ l PBS is loaded into an activated well of a module and subjected to microwave irradiation for 10 seconds inside a domestic microwave oven (BPL-Sanyo, India), operating at a frequency of around 2450 MHz with a maximum power output of around 700 watts. Irradiation is conducted in the microwave oven with the highest power setting of 10, that is the magnetron duty cycle of 100% of an output power of 700 watts.

The well is thoroughly washed with washing buffer so as to remove the unbound antigen. Subsequent steps are carried out by conventional procedure. Thus, blocking with blocking solution (200 μ l), antibody (100 μ l) binding and anti rabbit IgG-horse radish peroxidase conjugate (100 μ l) binding are carried out by incubation at 37°C for 2 hours each. The well is washed thoroughly after each step by washing buffer. Color development is done using 100 μ l of substrate solution. The well is read at 490 nm in an ELISA Reader (Spectramax 190 microplate spectrophotometer, Molecular Devices Corporation, California 94089) and absorbance values are recorded. All the experiments are performed in triplicate wells. Similar experiments are conducted with -ve sera.

A control experiment using microwaves is carried out in a similar manner using untreated wells. Another control experiment is performed using the activated wells by incubating the antigen at 37°C for the same duration as is done under microwaves. There is no immobilization of antigen in both the control reactions.

The whole experiment is separately repeated by varying the time for antigen binding viz. 30, 50, 70, and 90 seconds each.

The results for optimization of antigen binding time for microwave irradiation are given in Table-1

EXAMPLE 3**Blocking of free surface with blocking agent by microwave irradiation.**

E. histolytica antigen is immobilized by microwaves in an activated well of a module in 70 seconds as above. After thorough washing with washing buffer 200 µl of blocking solution is added to the well and irradiated with microwaves at 700 watts for 10 seconds. Subsequent steps of antibody and conjugate binding are done outside microwave oven as described in example 2. Color development and absorbance reading are also done as in example 2.

Similar experiments are conducted with -ve sera.

To check the optimum time for blocking, two different experiments are carried out in a similar manner except the microwave exposure time is increased to 40 and 60 seconds. All the experiments are performed in triplicate wells.

The results for optimization of blocking time under microwave irradiation are given in Table-2.

EXAMPLE 4**Antibody binding by microwave irradiation at high energy level.**

E. histolytica antigen is immobilized onto the activated well of a module by microwaves at 700 watts in 70 seconds followed by blocking of free surface with blocking agent by microwave irradiation at 700 watts for 10 seconds as in example 3. Antiamoebic antibody (100 µl) is loaded into the well. The well is then exposed to microwaves for 10 seconds at 700 watt. After washing the wells properly with washing buffer, binding of conjugate and subsequent color development are carried out as in example 3. Similar experiments are conducted with -ve sera. The experiment is repeated varying the time of microwave exposure at 30, 50, 70 and 90 seconds for binding antibodies, keeping all other conditions similar. All the experiments are performed in triplicate wells.

The results for antibody binding by microwave irradiation at high energy level are shown in Table-3.

EXAMPLE 5

Antibody binding by microwave irradiation at low energy level.

5 The experiments are performed here for antibody binding in the similar conditions as described in example 4, except that the antibody binding step is performed at a low energy level that is at 155 watts. Time of microwave exposure is 10, 50, 100 and 150 seconds respectively for four different sets of experiments.

10 The results for antibody binding by microwave irradiation at low energy level are shown in Table-4.

EXAMPLE 6

Conjugate binding by microwave irradiation at high energy level.

15 *E. histolytica* antigen is immobilized onto an activated well by microwaves at 700 watts in 70 seconds followed by blocking by microwaves at 700 watts for 10 seconds and antibody binding at 155 watts in 100 seconds as in example 5.

100 µl of anti rabbit IgG-horse radish peroxidase conjugate is loaded into the well and is subjected to microwave irradiation at 700 watts. Time of microwave exposure is 5,10, 15 and 20 seconds respectively for four different sets of experiments keeping all other conditions similar as in example 5. All the experiments are performed in triplicate wells.

20 The results for conjugate binding by microwave irradiation at high energy level are shown in Table-5.

EXAMPLE 7

Conjugate binding by microwave irradiation at low energy level.

25 The experiments are performed here for second antibody-conjugate binding in the similar conditions as described in example 6, except that the second antibody-conjugate

binding step is performed at a low energy level that is at 155 watts. Time of microwave exposure is kept at 50, 100 and 120 seconds respectively for four different sets of experiments.

The results for conjugate binding by microwave irradiation at low energy level are shown in Table-6.

EXAMPLE 8

Detection of *E. histolytica* antibodies by Microwave mediated Enzyme-Linked Immunosorbent Assay (MELISA) and Enzyme-Linked Immunosorbent Assay (ELISA)

E. histolytica antigen is immobilized onto the activated well by microwaves at 700 watts in 70 seconds, blocked with blocking solution by microwaves at 700 watts for 10 seconds, antibody binding at 155 watts in 100 seconds followed by antibody-conjugate binding at 155 watts in 100 seconds as described in above examples. Similar experiments are conducted with -ve sera. All the experiments are performed in duplicate wells and repeated for 5 times. After each step thorough washing are done by washing buffer.

Conventional ELISA is performed with same reagents, substrate and buffer except that all the steps are carried out without microwave stimulation. Thus, ELISA is carried out by coating the activated wells with antigen overnight at 4°C, followed by blocking, antibody and conjugate binding at 37°C for 2 h each. Color development is same for the invented and the conventional procedure.

The results for the detection of *E. histolytic* antibodies by Microwave mediated Enzyme-Linked Immunosorbent Assay (MELISA) and Enzyme-Linked Immunosorbent Assay (ELISA) are presented in the Table-7

EXAMPLE 9

Detection of *A. fumigatus* antibodies by Microwave mediated Enzyme-Linked Immunosorbent Assay (MELISA) and Enzyme-Linked Immunosorbent Assay (ELISA)

Detection of *A. fumigatus* antibodies in patients sera is carried out by Microwave mediated Enzyme-Linked Immunosorbent Assay (MELISA) and Enzyme-Linked Immunosorbent Assay (ELISA) method in a similar conditions as described in example 8, except that the antigen is *A. fumigatus*, antibody is from 10 different patients sera, control sera having no specific antibody is from healthy volunteers and the conjugate is anti human IgG-peroxidase. All the experiments are performed in duplicate wells.

The results for the detection of *A. fumigatus* antibodies by Microwave mediated Enzyme-Linked Immunosorbent Assay (MELISA) and Enzyme-Linked Immunosorbent Assay (ELISA) are presented in the Table-8

Table 1. Detection of *E. histolytica* antibodies by carrying out first step by MELISA and remaining steps by ELISA procedure.

MELISA: Step-1. Immobilization of antigen by microwave irradiation at 700 watts onto the activated wells in different times as in the table, control: 37°C, 70 seconds

ELISA: (Step - 2 to 5) Conventional procedure

Time (in sec)	+ve sera			- ve sera			Remarks
10	0.188	0.196	0.191	0.002	0.004	0.003	++
30	0.208	0.193	0.198	0.005	0.001	0.002	++
50	0.226	0.211	0.214	0.006	0.002	0.004	+++
70	0.363	0.355	0.358	0.005	0.001	0.003	++++
90	0.370	0.360	0.367	0.001	0.003	0.004	++++
Control	0.003	0.001	0.004	0.006	0.004	0.002	-

Table 2: Detection of *E. histolytica* antibodies by carrying out first two steps by MELISA and remaining steps by ELISA procedure.

MELISA: Step-1. Ag binding- 70 seconds, 700 watts. Step-2. Blocking- variable time as in

the table, 700 watts. ELISA: (Step - 3 to 5) Conventional procedure.

Time (in sec)	+ve sera			- ve sera			Remarks
10	0.270	0.288	0.273	0.006	0.005	0.008	++++
40	0.352	0.377	0.367	0.098	0.092	0.094	++
60	0.293	0.290	0.287	0.283	0.270	0.276	-

Table 3: Detection of *E. histolytica* antibodies by carrying out first three steps by MELISA and remaining steps by ELISA procedure.

- 5 MELISA: Step-1. Ag binding- 70 seconds, 700 watts. Step-2. Blocking- 10 seconds, 700 watts. Step-3. Antibody binding-variable time as in the table, 700 watts. ELISA: (Step - 4 & 5) Conventional procedure.

Time(in sec)	+ve sera			- ve sera			Remarks
10	0.216	0.203	0.211	0.087	0.123	0.098	+
30	0.392	0.353	0.388	0.100	0.137	0.114	-
50	1.413	1.177	1.213	0.194	0.180	0.194	++
70	1.313	1.253	1.276	1.263	1.279	1.275	-
90	1.287	1.239	1.238	1.288	1.248	1.234	-

- 10 **Table 4:** Detection of *E. histolytica* antibodies by carrying out first three steps by MELISA and remaining steps by ELISA procedure.

MELISA: Step-1. Ag binding- 70 seconds, 700 watts. Step-2. Blocking- 10 seconds, 700 watts. Step-3. Antibody binding-variable time as in the table, 155 watts. ELISA (Step - 4 to 5): Conventional procedure.

Time (in sec)	+ve sera			- ve sera			Remarks
10	0.010	0.014	0.010	0.004	0.003	0.007	-
50	0.079	0.087	0.82	0.003	0.006	0.005	+
100	0.511	0.524	0.530	0.014	0.027	0.020	++++
150	0.256	0.276	0.289	0.218	0.221	0.223	-

Table 5: Detection of *E. histolytica* antibodies by carrying out first four steps by MELISA and last step by conventional procedure.

MELISA: Step-1. Ag binding- 70 seconds, 700 watts. Step-2. Blocking- 10 seconds, 700 watts. Step-3. Antibody binding- 100 seconds, 155 watts. Step-4. Conjugate binding- variable
5 time as in the table, 700 watts. Step - 5. Color development- 5 minutes at room temperature.

Time (in sec)	+ve sera			- ve sera			Remarks
5	0.147	0.136	0.138	0.032	0.020	0.23	++
10	0.166	0.166	0.168	0.087	0.093	0.85	+
15	0.192	0.171	0.183	0.110	0.119	0.116	-
20	0.388	0.503	0.446	0.312	0.288	0.293	-

Table 6: Detection of *E. histolytica* antibodies by carrying out first four steps by MELISA and last step by conventional procedure.

MELISA: Step-1. Ag binding- 70 seconds, 700 watts. Step-2. Blocking- 10 seconds, 700
10 watts. Step-3. Antibody binding- 100 seconds, 155 watts. Step-4. Conjugate binding- variable time as in the table, 155 watts. Step - 5. Color development- 5 minutes at room temperature.

Time (in sec)	+ve sera			- ve sera			Remarks
50	0.205	0.189	0.192	0.021	0.018	0.022	++
100	0.558	0.532	0.543	0.036	0.031	0.032	++++
120	0.145	0.193	0.157	0.023	0.014	0.023	+

Table 7: Detection of *E. histolytica* antibodies: Comparison of MELISA and ELISA procedures.

15 MELISA: Step-1. Ag binding- 70 seconds, 700 watts. Step-2. Blocking- 10 seconds, 700 watts. Step-3. Antibody binding- 100 seconds, 155 watts. Step-4. Conjugate binding- 100 seconds, 155 watts. Step - 5. Color development- 5 minutes at room temperature.

ELISA: Step-1. Ag binding- overnight at 4°C. Step-2. Blocking- 2h at 37°C. Step-3. Antibody binding- 2h at 37°C Step-4. Conjugate binding- 2h at 37°C Step - 5. Color

development- 5 minutes at room temperature.

Serial No.	+ve sera				-ve sera			
	MELISA		ELISA		MELISA		ELISA	
1	0.567	0.556	0.539	0.560	0.036	0.070	0.046	0.040
2	0.531	0.556	0.538	0.534	0.096	0.056	0.064	0.070
3	0.541	0.563	0.549	0.547	0.098	0.053	0.064	0.050
4	0.558	0.545	0.574	0.544	0.052	0.080	0.074	0.110
5	0.557	0.564	0.558	0.540	0.049	0.079	0.056	0.090
Remarks	Reproducible		Reproducible		Reproducible		Reproducible	

Table 8: Detection of *Aspergillus fumigatus* antibodies: Comparison of MELISA and ELISA procedures. Same procedures are followed as described in Table-7.

Sample No.	+ve sera				-ve sera			
	MELISA		ELISA		MELISA		ELISA	
1	0.334	0.356	0.339	0.320	0.036	0.070	0.116	0.130
2	0.331	0.356	0.438	0.380	0.096	0.056	0.094	0.110
3	0.351	0.380	0.379	0.410	0.098	0.053	0.164	0.150
4	0.498	0.525	0.574	0.520	0.052	0.080	0.174	0.170
5	0.685	0.664	0.658	0.641	0.049	0.079	0.056	0.080
6	0.367	0.375	0.372	0.390	0.063	0.047	0.093	0.110
7	0.430	0.449	0.412	0.400	0.114	0.131	0.110	0.150
8	0.550	0.560	0.527	0.510	0.090	0.077	0.098	0.100
9	2.204	2.081	1.984	2.300	0.058	0.061	0.073	0.120
10	0.346	0.341	0.327	0.310	0.039	0.037	0.142	0.160
Remarks	Comparable				Less non specific binding in MELISA			

5

Apparatus for MELISA

The apparatus for MELISA can be constructed with the following components:

(a) Loading chamber: It is the chamber for loading the samples or reagents from a specified bottle through a fine tube and a suitable pump onto the activated polystyrene plate/module

automatically.

(b) Reaction chamber: Reaction chamber is consisting of magnetron, exhaust fan and a light focus for carrying out the steps of binding of the antigen, blocking, antibody binding and antibody enzyme conjugate binding by microwave irradiation and enzyme substrate reaction at ambient temperature in a pre-programmed time as claimed in claim 1.

(c) Washing cum drying chamber: In this chamber washing and drying of the ELISA plate or module is done automatically by a pre-programmed command after each step of the MELISA procedure ;

(d) Detection chamber: This chamber is used for colorimetric detection with the help of the spectrophotometer;

(e) Moving platform: It is used for carrying the Elisa plate/modules from one chamber to another chamber;

(f) Control unit: It has microprocessor based computing means for controlling MELISA method as claimed in claim 1 through suitable hardware and software.

Advantages of the invention

Conventional methods of ELISA usually take several hours to 2 days for completion, which is the major drawback for a procedure used worldwide in different fields apart from clinical diagnostics. In case of medical urgency precious time is lost in diagnosis before the patient could be given medication. Therefore, a rapid ELISA procedure invented herein (MELISA) will be beneficial and useful for diagnosis of diseases, biomedical research and other related fields. Main advantages of the invented ELISA procedure are:

1. The invented procedure is very fast than the existing methods of ELISA.
2. The total time required in the invented method is less than 10 minutes. Thus it obviates the time consuming cumbersome procedure.

3. The invented procedure is very sensitive and requires minute quantities of precious

antigen or antibody.

4. The invented procedure is very accurate as the enzyme –substrate reaction is done in solution and quantified spectrophotometrically.
5. The procedure is simple and does not require any additional expertise or reagent to do it.
- 5 6. The invented procedure is cost effective and does not require any additional equipment except a domestic microwave oven, which is common in most of the laboratories.
7. The invented procedure is reproducible which is an important criterion for ELISA.
8. The procedure gives minimal or negligible non-specific binding.
9. The procedure has the potential for automation, which can minimize human error, which
10 usually varies from person to person.
10. The invented procedure has the potential for application in other immunoassays like radio immunoassay, radio-immunosorbent test, radio allergosorbent test, biotin- avidin /streptavidin immunoassay, immunoblotting, immunostaining etc. apart from different types of ELISA such as direct ELISA, indirect ELISA, sandwich ELISA and alike.

15 Thus the invented ELISA procedure which is rapid, economical, reproducible, simple and has a potential for automation. It will be beneficial to human kind due to its increasing importance in clinical diagnostics, molecular biology, agriculture, food technology, environmental science, biomedical research and other related fields

20

25

References.

1. Douillard, J.Y. and Hoffman, T. (1983) *Methods in Enzymology* **92**, 168-174.
2. Van Emon, J.M. and Lopez-Avila, V. (1992) *Analytical Chemistry*, **64**, 79 A-88A.
3. Linde, D.G. and Goh, K.S.(1995) *Pesticide Outlook*,18-23
- 5 4. Salgame, P., Varadhachary, A.S, Primiano, L.L., Finke, J.E., Muller, S. and Monestier, M. (1996) *Nucleic Acids Res.***25**, 680-681.
5. Satoh, A., Fukui, S., Yoshino, S. Shinoda, M., Kozima, K. and Matsumoto, I. (1999) *Analytical Biochemistry* **275**, 231-235.
6. Larsson, P.H., Johansson, S.G.O., Hult, A. and Gothe, S. (1987) *Journal of*
10 *Immunological Methods* **98**, 129-135.
7. Boon, M.E and Kok, L.K. (1992) Microwave irradiation in immunostaining, p. 256-285.
In *Microwave Cookbook of Pathology: The Art of Microscopic Visualisation*. 3rd. Ed.
Columbia press Leyden, Leiden.
8. Boon, M.E., Kok, L.K., Moorlag, H.E. and Suurmeijer (1989). *Am.J.Clin. Pathol.* **92**:137-
15 143.
9. Chiu, K.Y. and K.W. Chan. 1987. *J.Clin.Pathol.* **40**:689-692.
10. Hjerpe, A., Boon, M.E. and Kok, L.P. (1988). *Histochem.J.* **20**:388-396.
11. Koh, L.P. and Boon, M.E. (1992) *Microwave Cookbook for Microscopists: Art and*
Science of Visualization, Third Edition. Coulomb Press Leyden: The Netherlands
- 20 12. Sawhney, S., Chakravarti, R.N., Jain, P. and Vinayak, V.K. 1980,
Trans.Roy.Soc.Trop.Med.Hyg. **74**: 26-29
13. Sharma, G.L, Naik, S.R. and Vinayak, V.K. 1984, *Aust.J.Exp.Biol.Med.Sc.* **62**: 117-133.
14. Lowry, O.H., Rosebrough, N.J., Farr, A.L. and Randall, R.J. 1951, *J.Biol.Chem.* **193**:
265.
- 25 15. Voller A., Bidwell D, Bartett A. Microplate ELISA and its application, In

Immunoenzymatic Assay Techniques. Malvano R. ed. The Hague Martinus Nijhoff Publ. 1980, p 104-115.

16. Banerjee, B., Chetty, A., Joshi, A.P., and Sarma, P.U., 1990, *Asian Pacific J Allergy immunol.* 8:13-18).
- 5 17. Rosenberg, M., Patterson, R., Mintzer, R., Cooper, B.J., Roberts, M. and Harris, K.E., *Ann Intern Med* 1997, 86:405-414).

Other References

10

Patent Documents

Apr., 1989

Boon *et al*

PCT patent application WO 89/ 03038

15

20

25

CLAIMS

1. A rapid method for microwave mediated enzyme-linked immunosorbent assay characterized in using an activated solid support wherein the said method comprises:
- (a) providing an activated solid support,
 - 5 (b) loading a biomolecule selected from an antigen or antibody by dissolving the said biomolecule in a coating buffer into the activated well of the said solid support and placing the said well inside a microwave oven followed by irradiating the said well with microwaves at a frequency ranging between 2300-2500 MHz with the power output ranging between 600-900 watts for a period ranging from 50-100 seconds followed by
10 washing the well thoroughly with an appropriate washing buffer,
 - (c) blocking the free sites of the well with an immobilized biomolecule as obtained from step (b) as above by loading blocking solution into the said well and irradiating it inside the microwave oven at a frequency of from 2300-2500 MHz with a power out put ranging between 600-800 watts for a period ranging from 5-20 seconds and washing the said
15 well with an appropriate washing buffer,
 - (d) loading the corresponding antibody or antigen dissolved in a buffer into the well immobilized with antigen or antibody as obtained from step (c) above followed by irradiation of said well inside the microwave oven at a frequency of from 2300-2500 MHz with a power output ranging from 50-200 watts for a period ranging from 90-200
20 seconds followed by washing with washing buffer,
 - (e) loading an appropriate enzyme- conjugate dissolved in a suitable buffer into the above said well obtained from step (d) and irradiating the said well inside a microwave oven at a frequency of from 2300-2500 MHz with a power output ranging from 100-300 watts for a period ranging from 50-150 seconds followed by washing with a washing buffer,
 - 25 (f) adding a substrate-dye-buffer to the above well as obtained from step (e) as above and

keeping it for a period ranging from 4 to 10 minutes in dark followed by addition of stop solution and measuring optical density of the solution by spectrophotometer at a suitable wavelength.

2. A method as claimed in claim 1 wherein the solid support used is selected from the group consisting of polystyrene, polypropylene, polyethylene, glass, cellulose, nitrocellulose, silicagel, polyvinyl chloride, polyaniline and alike.
3. A method as claimed in claim 1 wherein the preferred solid support used is polystyrene.
4. A method as claimed in claim 1 wherein the solid support is selected from any shape, form and size such as sheets, plates, test particles such as beads and microspheres, test tubes, test sticks, test strips, well, ELISA plate, microwell plate or module.
5. A method as claimed in claim 1 wherein the solid support used for immobilizing biomolecules is selected from any support having atleast one active functional group selected from the group consisting of halide, aldehyde, acetyl, epoxide, succinamide, isothiocyanate, acylazide and alike.
6. A method as claimed in claim 1 wherein the functional group may be present in the support itself or can be introduced by conventional chemical or photochemical or other methods known to prior art.
7. A method as claimed in claim 1 wherein antigen either elicits or has the potential to elicit an immune response.
8. A method as claimed in claim 1 wherein microwave irradiation is performed in an apparatus or chamber where microwave can be generated and is selected from domestic microwave oven, specially designed microwave oven or any apparatus or chamber in which microwave is generated.
9. A method as claimed in claim 1 wherein total time for antigen binding, blocking, antibody binding and conjugate binding is ranging from 195 to 470 seconds.

10. A method as claimed in claim 1 wherein blocking agent is selected from the group
consists of bovine serum albumin, skimmed milk powder and gelatin.
11. A method as claimed in claim 1 wherein coating buffer is selected from carbonate buffer,
phosphate buffer having a pH, in the range of from 6.5 to 11, with molarity ranging from
5 0.005 M to 0.1 M.
12. A method as claimed in claim 1 wherein washing buffer used is a mixture of phosphate
buffer saline having a pH, in the range of from 6.5 to 11, with molarity ranging from
0.005 M to 0.1 M and tween 20 in the range between 0.05% to 3%.
13. A method as claimed in claim 1 wherein conjugate used is selected from biomolecule
10 having antibody or antigen conjugated with an enzyme selected from peroxidase or
alkaline phosphatase.
14. A method as claimed in claim 1 useful for carrying out assays selected from the group
consisting of radio immunoassay, radio-immunosorbent test, radio allergosorbent test,
biotin- avidin /streptavidin immunoassay, immunoblotting, immunostaining etc. apart
15 from different types of ELISA such as direct ELISA, indirect ELISA, sandwich ELISA
and alike.
15. An apparatus for microwave mediated enzyme linked immunosorbent assay (MELISA)
comprising
- (a) a loading chamber, for loading the samples or reagents from a specified bottle through
20 a fine tube and a suitable pump onto the activated polystyrene plate/module
automatically,
- (b) a reaction chamber consisting of magnetron, exhaust fan and a light focus for carrying
out the steps of binding of the antigen, blocking, antibody binding and antibody enzyme
conjugate binding by microwave irradiation and enzyme substrate reaction at ambient
25 temperature in a pre-programmed time as claimed in claim 1.

(c) a washing cum drying chamber for washing and drying the said ELISA plate or module automatically by a pre-programmed command after each step of the MELISA procedure ;

(d) a detection chamber for colorimetric detection with the help of the spectrophotometer ;

(e) a moving platform is used for carrying the Elisa plate/modules from one chamber to another chamber ;

(f) microprocessor based computing means for controlling MELISA method as claimed in claim 1 through suitable hardware and software.

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 G01N33/543

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

BIOSIS, EPO-Internal, PAJ, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>BUDDE U ET AL: "DRASTISCHE VERKUEZUNG VON INKUBATIONSZEITEN BEI IMMUNHAEMATOLOGISCHEN UNTERSUCHUNGEN DURCH EINSATZ VON MIKROWELLENGERAETEN" INFUSIONSTHERAPIE UND TRANSFUSIONSMEDIZIN, BASEL, CH, vol. 22, no. SUPPL. 01, 1995, pages 92-94, XP000985251 ISSN: 1019-8466 the whole document</p> <p style="text-align: center;">--- -/-</p>	1-14

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- *&* document member of the same patent family

Date of the actual completion of the international search

10 May 2001

Date of mailing of the international search report

22/05/2001

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Stricker, J-E

INTERNATIONAL SEARCH REPORT

International Application No

PCT/IN 00/00075

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	ZHANG L-Z ET AL: "USE OF MICROWAVES IN IMMUNOENZYME TECHNIQUES" CLINICAL CHEMISTRY, AMERICAN ASSOCIATION FOR CLINICAL CHEMISTRY. WINSTON, US, vol. 39, no. 9, September 1993 (1993-09), page 2021 XP000985131 ISSN: 0009-9147 abstract Paragraph "Materials and Methods"	1-13
Y	MARANI ENRICO: "Microwave applications in neuromorphology and neurochemistry: Safety precautions and techniques." METHODS (ORLANDO), vol. 15, no. 2, June 1998 (1998-06), pages 87-99, XP002166951 ISSN: 1046-2023 abstract page 92, column 2, paragraph 5 Chapter "Neurochemistry" on pages 94-95	1-14
Y	DORP VAN R ET AL: "ELISA INCUBATION TIMES CAN BE REDUCED BY 2.45-GHZ MICROWAVES" JOURNAL OF CLINICAL AND LABORATORY IMMUNOLOGY, TREVIOT-KIMPTON PUBLICATIONS, LONDON, GB, vol. 34, no. 2, February 1991 (1991-02), pages 87-96, XP000985204 ISSN: 0141-2760 abstract Chapters "Materials and methods" and "Discussion" page 95, column 1, paragraph 3 page 95, column 2, paragraph 4	1-13
Y	DORP VAN R ET AL: "A RAPID ELISA FOR MEASUREMENT OF ANTI-GLOMERULAR BASEMENT MEMBRANE ANTIBODIES USING MICROWAVES" JOURNAL OF CLINICAL AND LABORATORY IMMUNOLOGY, TREVIOT-KIMPTON PUBLICATIONS, LONDON, GB, vol. 40, no. 3, 1993, pages 135-147, XP000985202 ISSN: 0141-2760 abstract Chapters "Materials and methods" and "Results" page 146, column 1, paragraph 3 --- -/-	1-13

INTERNATIONAL SEARCH REPORT

Int. Application No.

PCT/IN 00/00075

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 89 03038 A (BOON MATHILDE ELISABETH ;KOK LANBRECHT PIET (NL)) 6 April 1989 (1989-04-06) cited in the application	15
A	page 3, paragraph 1 claims 1,2	1-14
Y	----- PATENT ABSTRACTS OF JAPAN vol. 1996, no. 07, 31 July 1996 (1996-07-31) & JP 08 082627 A (SUZUKI MOTOR CORP), 26 March 1996 (1996-03-26) abstract	15
Y	----- EP 0 604 970 A (CEM CORP) 6 July 1994 (1994-07-06) abstract figure 1	15
Y	----- US 5 304 766 A (BAUDET JEAN-JACQUES ET AL) 19 April 1994 (1994-04-19) abstract figure 1	15
Y	----- US 5 455 008 A (EARLEY JAMES J ET AL) 3 October 1995 (1995-10-03) abstract column 11, line 11-18 claims 1,9,22 figures 1-9 -----	15

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/JP 00/00075

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 8903038 A	06-04-1989	NL 8702338 A	17-04-1989
JP 08082627 A	26-03-1996	NONE	
EP 0604970 A	06-07-1994	US 5420039 A	30-05-1995
		CA 2111383 A	01-07-1994
		JP 6292885 A	21-10-1994
US 5304766 A	19-04-1994	FR 2681431 A	19-03-1993
		AT 172027 T	15-10-1998
		AU 649770 B	02-06-1994
		AU 1030792 A	30-07-1992
		CA 2060037 A,C	26-07-1992
		DE 69227208 D	12-11-1998
		DE 69227208 T	27-05-1999
		EP 0496684 A	29-07-1992
		ES 2124722 T	16-02-1999
		JP 2843702 B	06-01-1999
		JP 5190277 A	30-07-1993
		KR 219888 B	01-09-1999
US 5455008 A	03-10-1995	WO 9408759 A	28-04-1994